CLINICAL IMPLICATIONS OF THE MOLECULAR BIOLOGY OF PROSTATE CANCER: REVIEW ARTICLE

IMPLICANCIAS CLÍNICAS DE LA BIOLOGÍA MOLECULAR DEL CÁNCER DE PRÓSTATA: ARTÍCULO DE REVISIÓN

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ABSTRACT

To understand the term genomic heterogeneity in prostate cancer, we must understand the clonal genomic evolution of cancer, as well as knowing that it is a dynamic and evolutionary phenomenon. Knowing the genome of prostate cancer not only allows us to have a vision over time of the genomic alterations that occur during its different stages, but also to learn about the mechanisms of metastasis. In addition, knowing the hereditary component of prostate cancer allows the evaluation of patients and to be able to identify if we are dealing with a family at risk.

Keywords: Prostatic Neoplasms; Genetic Heterogeneity; Germ-Line Mutation. (Source: MeSH - NLM)

RESUMEN

Para enteder el término de heterogeneidad genómica en cáncer de próstata debemos comprender la evolución genómica clonal del cáncer, así como saber que es un fenómeno dinámico y evolutivo. Conocer el genoma del cáncer de próstata no solo nos permite tener una visión en el tiempo de las alteraciones genómicas que se producen durante sus diferentes estadíos, sino también conocer sobre los mecanismos de la metástasis. Además, conocer el componente hereditario del cáncer de próstata permite la evaluación de los pacientes y poder identificar si estamos frente a una familia en riesgo.

Palabras claves: Neoplasias de la Próstata; Heterogeneidad Genética; Mutación de Línea Germinal.

INTRODUCTION

Prostate cancer is, from a molecular point of view, biologically heterogeneous due to the diversity of inter and intratumor molecular alterations since it is part of a dynamic and evolutionary genomic process. With the development of new molecular techniques, such as gene sequencing, microarrays, and epigenomic studies, among others, it has been possible to molecularly characterize prostate cancer, finding differences even between the different stages of the disease.

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In addition to molecular alterations at the somatic level in prostate tumor cells, there are also variants at the germ line level, that is, they are present in all the cells of your body from the moment of conception, giving them a higher risk of developing prostate cancer throughout life and at a younger age. Men who have a family history of prostate cancer, mainly first-degree, are more likely to develop the disease as it could be a hereditary syndrome.

The objective of this review article is to present molecular alterations at the somatic level in prostate cancer, its molecular classification, genomic heterogeneity, clonal evolution, biomarkers, as well as genetic predisposition syndromes to prostate cancer.

MOLECULAR ALTERATIONS AT THE SOMATIC LEVEL

The main molecular alterations described in prostate cancer include the TMPRSS2–ETS gene fusion, copy number variants of TP53, AR, RB1, PTEN/PIK3CA, BRCA2, and ATM genes, among others.

TMPRSS2-ETS gene fusion

TMPRSS2 (Transmembrane protease, serine two) is an androgen-regulated serine protease. ETS (erythroblastosis virus E26 oncogene homolog) is one of the largest families of transcription factors that includes ERG and ETV1, which are fused with the TMPRSS2 gene in approximately 50-79% of prostate cancer cases⁽¹⁾. This fusion is due to the deletion of a region between both genes and is associated with a worse prognosis in localized cases⁽²⁾.

PTEN/PI3K/AKT/mTOR pathway

The PTEN/PI3K/AKT signaling pathway plays a primordial role in the regulation of cell growth and death, while the PI3K/AKT/mTOR pathway plays a fundamental role in tumor metastasis ⁽³⁾. Variants in the PTEN and PI3K genes are mutually exclusive since the PTEN gene is a negative regulator of the PI3K/AKT pathway, so the loss of PTEN is associated with a worse prognosis ⁽¹⁾ two to 14% of prostate cancer cases

have variants in the PTEN gene and 12-41% of cases have copy number loss⁽⁴⁾. On the other hand, variants in the PI3K gene have been described in three to 4% of prostate cancer cases and its amplification has been reported in four to 10% of cases. In addition, cases of combined variants of PTEN and TP53 have been described, which are very aggressive⁽⁴⁾.

Tp53

The TP53 gene is a tumor suppressor that plays a very important role in maintaining genomic stability and preventing carcinogenesis. Approximately three to 47% of prostate cancer cases have variants in the TP53 gene and between two to 15% gene loss ⁽⁴⁾. Variants in the TP53 gene have been associated with an increased risk of disease recurrence⁽¹⁾.

AR

About two to 18% of prostate cancer cases have variants in the androgen receptor AR gene (androgen receptor gene) or gene amplification (five to 52% of cases), being very common in cases refractory to hormonal therapy⁽⁴⁾. Androgen receptors belong to the group of nuclear steroid hormone receptors and act as a liganddependent transcription factor, that controls the expression of specific genes⁽¹⁾.

RB

The RB protein is a product of the RB1 gene (Retinoblastoma one gene), a tumor suppressor which is mutated in approximately one to 4% of prostate cancer cases, while loss of the RB1 gene has been shown in five to 23%⁽⁴⁾. An alteration in the RB1 gene leads to a failure in the function of the protein, stimulating the release of the androgen receptor and conferring resistance to castration ⁽¹⁾. The RB1 gene is frequently deleted or methylated in castration-resistant prostate cancer⁽¹⁾.

APC

The APC gene is a tumor suppressor that encodes a protein that acts antagonistically to the Wnt signaling pathway ⁽¹⁾. Hypermethylation of the APC gene promoter is a predictor of poor prognosis in prostate cancer. Approximately three to 10% of prostate cancer cases have variants in the APC gene ⁽⁴⁾.

MYC

The MYC proto-oncogene encodes a protein that plays an important role in cell cycle progression, apoptosis, and transformation⁽¹⁾. Activation of this proto-oncogene transforms it into an oncogene that is expressed through gene amplification, thus stimulating cancer development, which is found in approximately two to 20% of prostate cancer cases⁽⁴⁾.

BRCA2

BRCA2 is a tumor suppressor gene, which is responsible



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for maintaining genomic stability, mainly in the doublestranded DNA repair pathway by homologous recombination⁽¹⁾. Approximately 9% of prostate cancer cases present variants in BRCA2, of which two to 6% are due to germline variants, which are a prognostic factor for survival in all stages of prostate cancer, including localized cases⁽⁵⁾.

When the variants are germinal, the relatives of the patient have a greater probability of having inherited the mutated gene, conferring risk of developing a neoplasm related to hereditary breast and ovarian cancer syndrome in carriers.

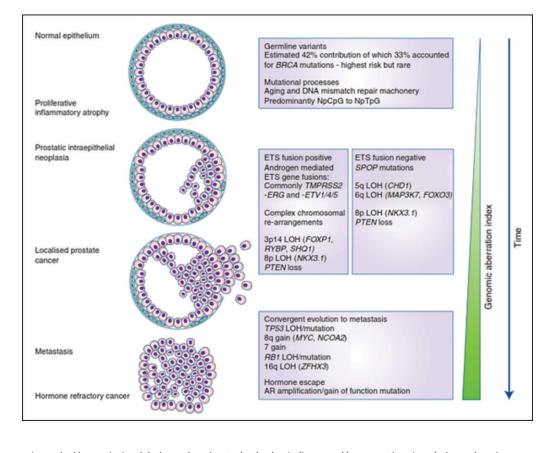
ATM

ATM is a DNA repair gene that acts at the cell cycle level as a necessary controller for the cellular response to DNA damage and to maintain genomic stability ⁽¹⁾. Approximately 5% of prostate cancer cases have somatic ATM variants, including 1% germline variants ⁽⁵⁾.

TMPRSS2-ETS GENE FUSION

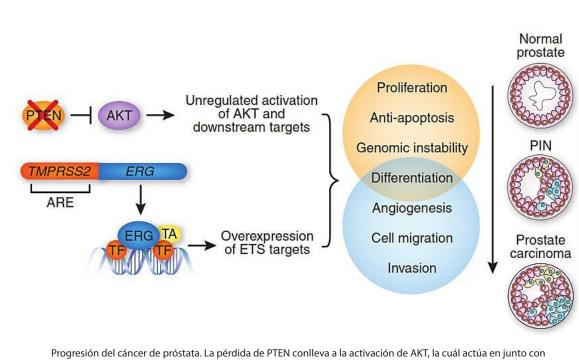
In 2005, Scott Tomlims et al. identified a recurrent rearrangement in more than half of the prostate cancer cases analyzed, which led to the gene fusion of the TMPRSS2 gene (21q22) with members of the ETS transcription factor family (ERG, ETV1, and ETV4 located at loci 21q22, 7p21 and 17q21, respectively)⁽⁶⁻⁸⁾.

Assessing the presence of the TMPRSS2-ETS gene fusion in prostate cancer samples serves as a prognostic marker of the disease. Determination of ERG overexpression by immunohistochemistry has been highly correlated with gene fusion status with 86% sensitivity and specificity⁽⁶⁾.

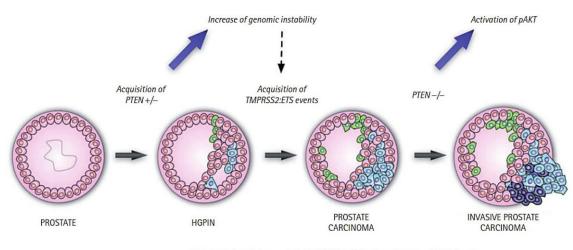


La evolución genómica del cáncer de próstata desde el epitelio normal hasta metástasis y el cáncer de próstata resistente a castración. Los estadíos patológicos del cáncer de próstata están representados en el lado izquierdo con la correspondiente mutación genómica en el lado derecho. Tomado de "The genomic evolution of human prostate cancer"⁽⁷⁾.





Progresión del cáncer de próstata. La pérdida de PTEN conlleva a la activación de AKT, la cuál actúa en junto con la proteína quimérica TMPRSS2-ERG para promover la progresión del cáncer de próstata, desde un estadío pre-maligno como la neoplasia prostática intraepitelial (PIN). ARE: androgen response elements; TA: transcriptional activator; TF: transcription factor. Tomado de "TMPRSS2-ERG and PTEN loss in prostate cancer" ^(a).



PTEN haplo-insufficiency ----- Complete loss of PTEN function

Modelo de la secuencia de eventos genómicos en la progresión del cáncer de próstata. La adquisición de la haploinsuficiencia de PTEN en precursores del cáncer de próstata puede ser un evento temprano en la caracinogénesis. La disminución de los niveles de la proteína PTEN puede facilitar la inestabilidad genómica llevando a la adquisición de rearreglos como TMPRSS2-ETS, actuando ambos de forma sinérgica en la progresión del cáncer de próstata. La continua inestabilidad conlleva a una heterogeneidad tumoral debido a la presencia de subclonas. La pérdida completa de la función de PTEN (homocigota) favorece a la progresión tumoral tras activar la vía de AKT⁽⁹⁾.

MOLECULAR CLASSIFICATION OF PROSTATE CANCER

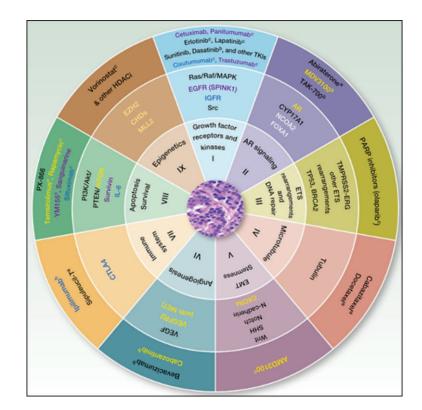
There is currently a molecular classification of primary prostate cancer. All seven molecular subtypes of primary prostate cancer, considered the earliest drivers of carcinogenesis, are defined by ERG fusions (46%), ETV1/ETV4/FL11 fusions or overexpression (8%, 4%, 1%, respectively), or by variants in the genes SPOP (11%), FOXA1 (3%) and IDH1 (1%) (nine to 11), while 26% is made up of other variants. Likewise, it has been determined that the molecular profiles in primary and metastatic prostate cancer are different.

(8)

GENOMIC HETEROGENEITY IN PROSTATE CANCER

The heterogeneity can be intratumor and intertumor, as well as in the different stages of prostate cancer, which is why we currently speak of molecular subtypes in prostate cancer. The high tumor heterogeneity has implications for the diagnosis, follow-up, and treatment of patients with prostate cancer ⁽¹²⁾.

Molecular studies such as next-generation gene sequencing, as well as studies of circulating tumor cells, epigenetic studies, and microarrays, among others, provide us with evidence of a clonal expansion of prostate cancer with the different mutational events that occur in its genome. The somatic level at various stages of disease progression. There are investigations of the genomic composition of multiple sites of metastasis in the same patient or have followed the clonal evolution longitudinally in the tissues. To understand the origin of prostate cancer, its behavior over time, and the risks of disease progression, it is necessary to know the spatial and temporal genomic evolution of prostate cancer.

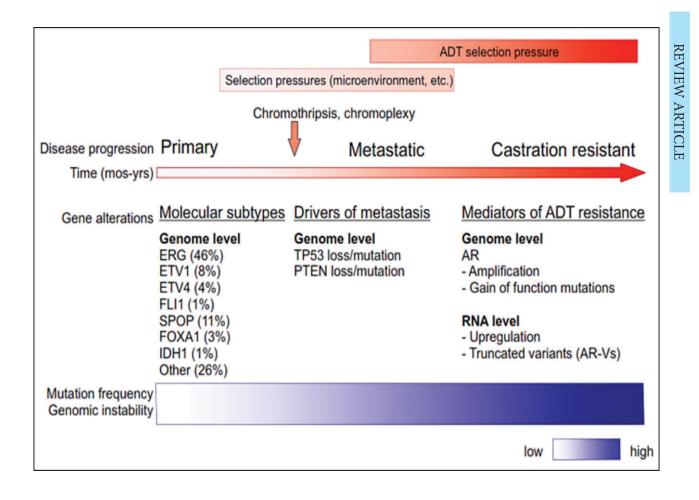


Terapia blanco basada en las vías moleculares alteradas en cáncer de próstata. Tomado de "Genomic Profiling Defines Subtypes of Prostate Cancer with the Potential for Therapeutic Stratification"⁽¹²⁾.

The clonal architecture of primary and metastatic tumors, allows us to know the clonal evolution during the progression of the disease ⁽¹³⁾. Various variants, as well as altered molecular pathways in both the primary tumor and metastasis, have been identified, confirming intratumor heterogeneity and thus evidencing that metastasis and resistance to androgen deprivation treatment occur due to different molecular alterations acquired over time⁽¹³⁾.

Studies reveal that the loss or variant of the TP53 gene, as well as the loss of PTEN, occur before or at the beginning of metastasis, indicating that they are drivers of metastatic spread ⁽¹³⁾. In addition to the seven molecular subtypes already mentioned (ETS fusions, variants in FOXA1, FLI1, SPOP, and IDH1) there may be an acquisition of new variants that lead to metastases, which could not be preferentially present in any of these subtypes⁽¹³⁾. Other important findings such as the androgen receptor (AR), which is altered in more than 60% of cases of metastatic prostate cancer, and which is also a mediator for resistance to androgen deprivation therapy (ADT), which can acquire new variants after metastasis⁽¹⁴⁾. What is not yet clearly known is whether these rare subclones originated in the primary tumor or early in the metastases, possess alterations in the AR that subsequently promote resistance to ADT, or whether these alterations arise after metastasis and initial ADT treatment ⁽¹³⁾.

The study of the prostate cancer genome not only allows us to have a vision over time of the genomic alterations that occur during the different stages of prostate cancer but also to learn about the mechanisms of metastasis. Metastatic spread can occur through monoclonal or polyclonal seeding between metastases, or in waves originating from the primary tumor ⁽¹⁴⁾. It has also been shown that clonal propagation is not only unidirectional, for example, when metastatic subclones reach the surgical bed of the resected primary tumor (figure 1 y 2).



Genes y vías moleculares alterados en diferentes estadíos del cáncer de próstata. Los factores ambientales imparten presiones de selección sobre las poblaciones clonales para conducir a la metástasis y en última instancia a la resistencia a la terapia de deprivación androgénica. La inactivación de TP53 y de PTEN se observan con más frecuencia en el cáncer de próstata resistente a castración (CRPC) metastásico en comparación con el cáncer de próstata primario, probablemente desempeñando un papel conductor de la metástasis. Los mediadores de la resistencia a la ADT se observan luego de la diseminación metastásica. Tomado de "Clonal origin and spread of metastatic prostate cancer" ⁽¹³⁾.

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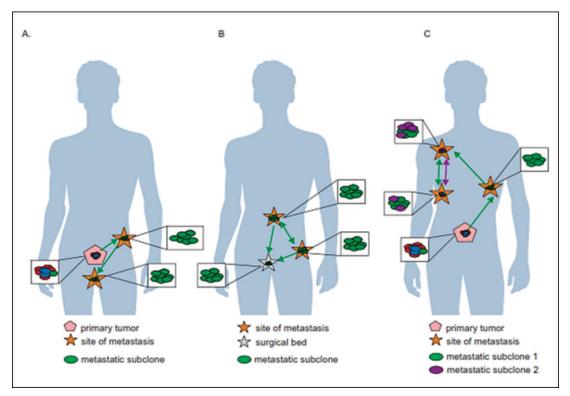


Figure 1. Clonal evolution of prostate cancer. A. A rare clone in the primary tumor (green) acquires metastatic potential and is seeded in different anatomical sites over time (green arrows). B. Metastatic subclones (green) that are seeded at different locations, including bidirectionally between metastatic sites (green arrows), can also seed the surgical bed again. C. The clonal populations of the different metastatic sites are planted in different anatomical sites but in a polyclonal way, such as metastatic subclone 1 (green) is planted in the left rib and right shoulder (green arrows), but a new subclone arises in the right shoulder (purple) (metastatic subclone 2), and both seed the right rib. Taken from "Clonal origin and spread of metastatic prostate cancer"⁽¹³⁾.

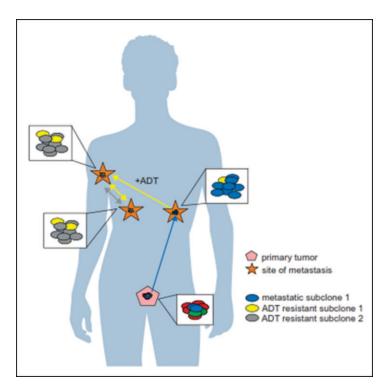
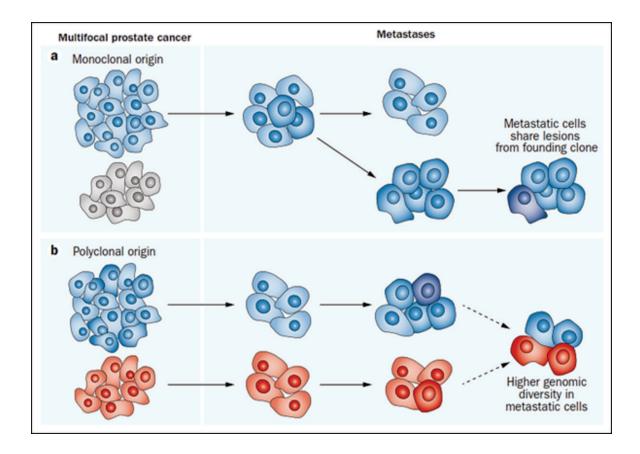


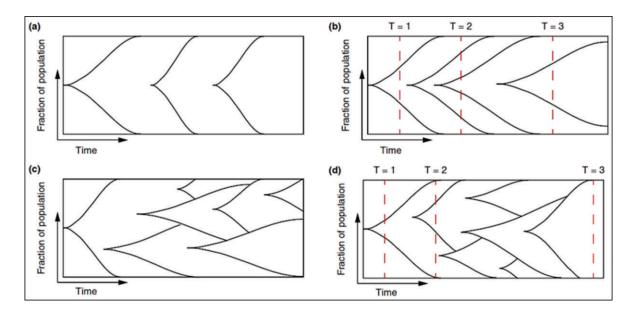
Figure 2. A rare clone in the primary tumor (blue) acquires metastatic potential (metastatic subclone 1) and is seeded elsewhere (blue arrow), where subclonal populations acquire new variants. Following androgen deprivation therapy (+ADT), a subclonal population (yellow) that possesses variants resistant to this treatment (ADT-resistant subclone 1) undergoes clonal expansion and spreads (yellow arrows) to a distant site. The acquisition of new variants can generate a new population of subclones resistant to ADT 2 (grey). The seed mix of clones resistant to therapy seed other sites in a polyclonal fashion, even bidirectionally (yellow and gray arrows). Taken from "Clonal origin and spread of metastatic prostate cancer" ⁽¹³⁾.



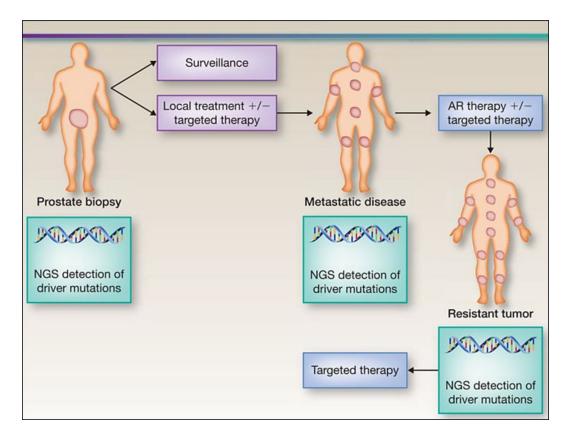
There is evidence that after radical prostatectomy, tumor samples have multiple intraprostatic cancer foci that are separate and may be biologically variable in their potential for aggressiveness and disease progression ⁽¹⁵⁾. Heterogeneity can be of two types, intratumor (different molecular alterations in multiple cancer foci in the tumor specimen from the same patient) and intertumor (such as different molecular alterations in patients with the same Gleason score) ⁽¹⁵⁾. Through next-generation gene sequencing, it has been possible to identify a variety of molecular alterations in prostate cancer such as gene fusions, amplifications, hemizygous and homozygous deletions, and/or point variants, which shows that there is genomic heterogeneity. Prostate cancer originates through a clonal evolutionary process, due to a sequential accumulation of variants to the point where these variants promote the development of a neoplastic phenotype (these variants are called "drivers")⁽¹⁶⁾, differs in its molecular composition at each stage of the disease. Some of these driver variants accelerate the acquisition of new variants by hindering the activity of some cellular mechanisms to detect or repair DNA damage or to respond with programmed cell death. The increase in the rate of variants leads to the acquisition of variants that have no immediate functional relevance for the cell (passenger variants) but that will later become relevant or even vulnerable as therapeutic targets ⁽¹⁷⁻¹⁹⁾.



Modelos monoclonales y policionales de metástasis de un cáncer de próstata primario multifocal. (a) En el modelo monoclonal, todas las lesiones metastásicas derivan de una célula antecesora en uno de los focos del cáncer de próstata primario multifocal. (b) En el modelo policional, múltiples focos genómicamente distintos en el cáncer primario de próstata pueden, independiente, progresar y hacer metástasis portando múltiples alteraciones clonales distintas a las originadas en el tumor primario. En ambos modelos, la adquisición subsecuente de mutaciones (azul y rojo oscuro) conlleva a una diversidad genómica. Tomado de "Intrapatient heterogeneity in prostate cancer" ⁽¹⁸⁾.



Trayectoria de la evolución del cáncer: lineal y ramificado. (a) Evolución lineal con extensiones clonales sucesivas, eliminando cada clon anterior. (b) Evolución lineal con extensiones clonales incompletas, donde se observa que en los puntos de tiempo T=2 y T=3 habría una heterogeneidad subclonal. (c) Evolución ramificada, donde surgen clonas hermanas de forma independiente y se expanden en paralelo creando una extensa heterogeneidad subclonal. En este tipo de evolución, las clonas hermanas compiten incluso por recursos como nutrientes y oxígenos, además de competir por el espacio (ambiente tumoral). (d) Pueden existir extensiones clonales en la evolución ramificada, lo que conlleva a la extinción de las ramas del árbol evolutivo que no eran ancestrales al clon recién dominante. Tomado de "The evolution of the unstable cancer genome"⁽¹⁹⁾.



Tomado de "New Strategies in Prostate Cancer: Translating Genomics into the Clinic"(17).

To perform precision medicine in patients with prostate cancer, it is necessary that, at the time of diagnosis through biopsy, molecular analysis is requested to detect conductive variants in the tumor, and based on its molecular profile, determine monitoring and treatment. Subsequently, if the disease progresses or if there is resistance to treatment, it is also necessary to carry out a molecular analysis, because the molecular profile is variable in each stage of the disease.

BIOMARKERS IN PROSTATE CANCER

Biomarkers are widely used for screening, detection, and prognosis of prostate cancer, revolutionizing the diagnosis and monitoring of the disease. Despite various studies on prostate cancer, there is no molecular biomarker with a potential clinical application for diagnosis and stratification in all stages of the disease, from localized to metastatic. Prostate-specific antigen (PSA) is usually used routinely as a marker, however, it has a low positive predictive value in localized prostate cancer and does not differentiate the stage of the disease ⁽²⁰⁻²¹⁾.

PCA3 (9q21-22) is a highly expressed non-coding mRNA in prostate tumors, which can be detected in urine and prostate fluid in patients with prostate cancer, in addition to being approved by the FDA and being available in various laboratories in the world. The cut-off point approved by the FDA is 25 since it is at this point that there is a balance between the sensitivity and specificity of the test ⁽²²⁾. Several studies also use the cutoff point 35 (22). A PCA3 value > 35 in urine has a sensitivity of 66% and specificity of 76% for the diagnosis of prostate cancer compared to PSA in serum (specificity 47% and sensitivity 65%)⁽²³⁾. The study of PCA3 in urine obtained after the prostate massage is superior to serum PSA in predicting the result of the biopsy with a sensitivity and specificity of 70% and 80% respectively, it also has a negative predictive value of 90% (22). The urine test can predict the presence of prostate cancer in a biopsy, with PCA3 levels being independent of prostate volume and serum PSA (23). Elevated levels of PCA3 have even been shown in

patients with elevated PSA and negative biopsies, which could help reduce the number of unnecessary biopsies ⁽²³⁾.

In addition to being a common rearrangement in prostate cancer, the TMPRSS2-ERG gene fusion serves as a molecular biomarker in prostate cancer with a specificity of 90% and a positive predictive value of 94% ⁽²⁴⁾. With the detection of this gene fusion, the probability of finding cancer in the biopsy increases from 15% to 90%, and even if the gene fusion is detected in urine and the biopsy does not detect cancer, it is necessary to repeat the biopsy, due to the high specificity of this biomarker (24). In addition, this gene fusion is associated with a poor prognosis in patients who carry it in the tumor⁽²³⁾. The combination of detection of TMPRSS2-ERG and PCA3 in urine improves the performance of PSA alone for prostate cancer detection and clinically meaningful cancer prediction (25)

Genetic alterations at the germinal level are molecular biomarkers, that allow the calculation of risk in patients with prostate cancer because it has been shown to influence the aggressiveness of prostate cancer and patient survival⁽²³⁾. In addition, the presence of variants in some of these genes is associated with the earlier presentation of cancer as well as a family history of prostate cancer.

Other promising biomarkers in prostate cancer are changes in DNA methylation, histone acetylation, or microRNAs, which can lead to gene silencing, resulting in altered gene expression without altering the DNA sequence ⁽²³⁾. Hypermethylation of the glutathione S-transferase P1 (GSTP1) gene is one of the most prevalent (90%) and can be detected in both serum and urine of patients; however, it is not specific for prostate cancer since it has been seen in approximately 70% of cases of high-grade intraepithelial neoplasia⁽²³⁾.

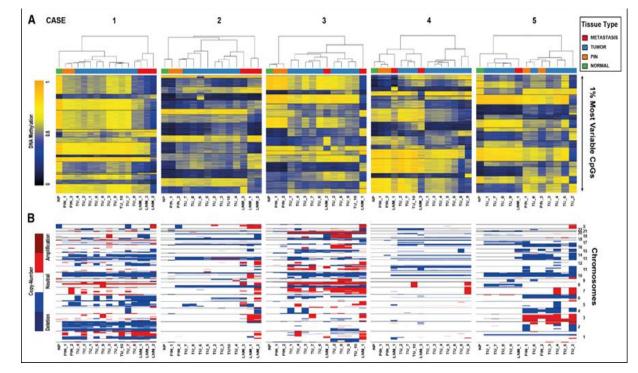
EPIGENETIC ALTERATIONS AND METHYLATION STUDIES IN PROSTATE CANCER

Changes in DNA methylation together with epigenetic silencing of some genes are the earliest somatic



changes identified in the development of prostate cancer ⁽²⁶⁾. The GSTP1 gene encodes an enzyme responsible for protecting the cell from genome damage⁽⁶⁾. The loss of GSTP1 expression is an early event in the onset of carcinogenesis since GSTP1 gene methylation has been shown in five to 10% of cases of

inflammatory proliferative atrophy and in more than 70% of prostatic intraepithelial neoplasia. high grade ⁽⁶⁾. GSTP1 promoter methylation is associated with a risk of recurrence in patients with prostate cancer, which makes it a marker of recurrence ⁽⁶⁾.



Poblaciones celulares subclonales difieren en sus perfiles genómicos y epigenómicos a nivel intratumoral mostrando la heterogeneidad en un mismo caso (casos del 1-5 de izquierda a derecha). (A) Tipo de tejido por cada caso está definido por colores: verde: tejido normal; naranja: lesión pre-maligna PIN; azul: tumor primario; rojo: metástasis ganglionar LNM. El nivel de metilación se presenta en rango, donde el color azul representa un bajo nivel de metilación y el color amarillo representa el mayor nivel de metilación (de 0% a 100%). (B) Alteraciones en el número de copias en diferentes regiones de un mismo tumor, donde las deleciones se representan en color azul y las amplificaciones en color rojo, las cuales se presentan de acuerdo a los cromosomas del 1 al X. Tomado de "Intratumor DNA Methylation Heterogeneity Reflects Clonal Evolution in Aggressive Prostate Cancer"⁽²⁶⁾.

CIRCULATING TUMOR CELLS IN PROSTATE

Among the new molecular biomarkers used in prostate cancer is the detection of circulating tumor cells (CTCs) in peripheral blood, and it is used as a prognostic tool since it has been proposed that the spread of CTCs in the blood is an essential mechanism of metastasis⁽²¹⁾.

Screening for CTCs is FDA-approved for monitoring the treatment of metastatic prostate cancer ⁽²¹⁾. In metastatic prostate cancer, the CTCs threshold per 7.5

ml of venous blood has been significantly determined as a prognostic marker for overall survival⁽²¹⁾.

GENETIC PREDISPOSITION TO PROSTATE CANCER

Prostate cancer has a hereditary component, which is characterized by the presence of cancer at an early age, having a family history of prostate cancer and other tumors, due to germline variants in BRCA1/2 genes, MMR (MLH1, MSH2, MSH6, PMS2), HOXB13, among others⁽²⁷⁾.

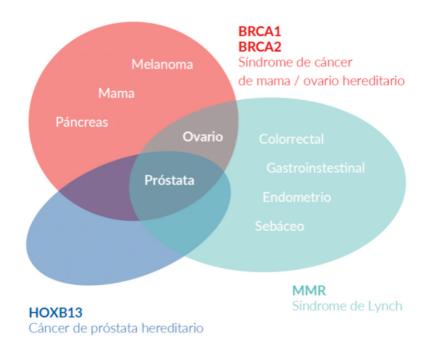


Figure 3. Genes and phenotypes of genetic syndromes with a predisposition to prostate cancer

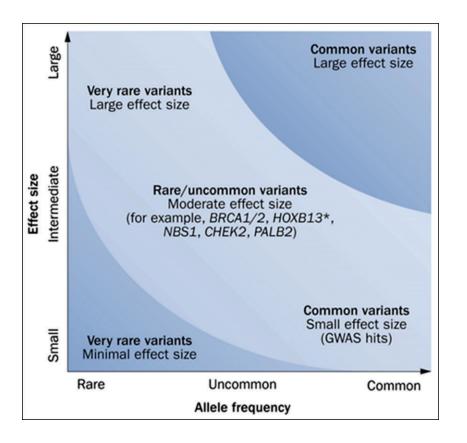


Figure 4. Model of frequent and rare genetic variants with a different effect on the genetic risk of developing prostate cancer. Taken from "Molecular Updates of Prostate Cancer" ⁽⁶⁾

 \cdot Age less than or equal to 65 years.

•Gleason > seven and family history of neoplasms related to Hereditary Breast Ovarian Cancer Syndrome.

•Family history of any cancer related to Hereditary Breast Ovarian Cancer Syndrome, hereditary prostate cancer, or Lynch syndrome, mainly in a first or seconddegree relative.

For the evaluation of patients with a suspected genetic predisposition to prostate cancer, it is necessary to carry out a family tree, to be able to identify if we are dealing with a family at risk. According to the types of neoplasms, age of presentation, and inheritance pattern, we will be able to define the genetic syndrome and thus direct the molecular study to identify germinal variants in the patient and their relatives.

The HOXB13 gene has been identified as a prostate cancer susceptibility gene⁽²⁸⁾. Germline variants in the HOXB13 gene have been associated with a two to eightfold increase in risk, these cases being those corresponding to hereditary prostate cancer⁽²⁷⁾. A germline variant in HOXB13, which is recurrent, G84E, p.(Gly84Glu), c.251G>A, has been frequently reported in patients diagnosed with prostate cancer at an early age (2.2%) and family history (3.1%)⁽²⁸⁾.

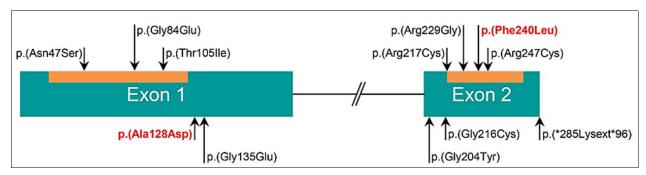


Figure 5. Structure of the HOXB13 gene and the distribution of pathogenic variants identified by gene sequencing reported in patients with prostate cancer. Taken from "Identification of Two Novel HOXB13 Germline Mutations in Portuguese Prostate Cancer Patients"⁽²⁸⁾

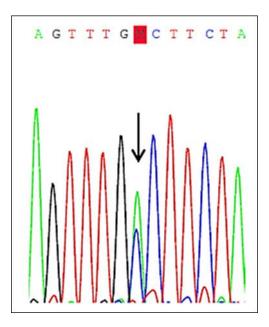
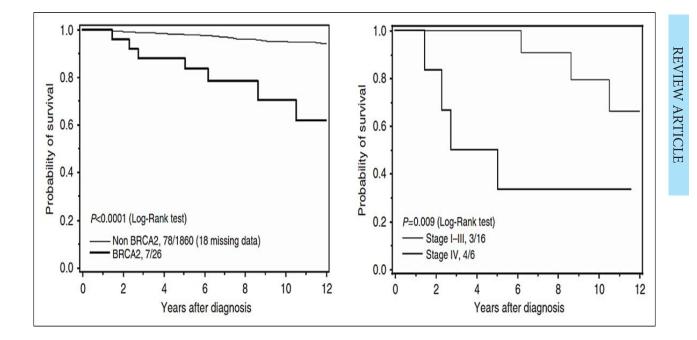


Figure 6. Sanger sequencing electropherogram showing the variant HOXB13 c.383C>A, p.(Ala128Asp) in heterozygosity at the germinal level. Taken from "Identification of Two Novel HOXB13 Germline Mutations in Portuguese Prostate Cancer Patients" ⁽²⁸⁾

Germline variants in the BRCA1 and BRCA2 genes correspond to cases of Hereditary Ovarian Breast Cancer Syndrome, where in addition to the risk of breast cancer, ovarian cancer, melanoma, and pancreas, there is a risk in men to develop prostate cancer ⁽²⁷⁾.

Germline variants in the BRCA2 gene have been associated with a more aggressive phenotype of the

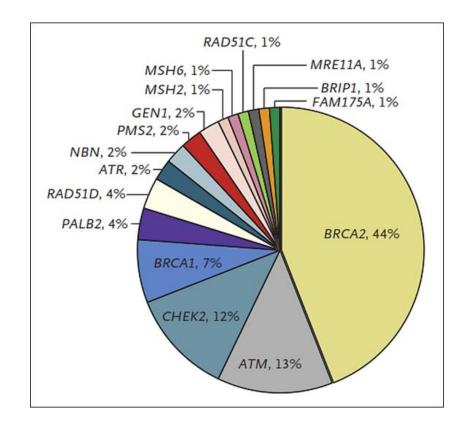
disease ⁽²⁹⁾. Likewise, germline variants in the BRCA2 gene confer a 7.3-8.6 times greater risk of developing prostate cancer before the age of 65, compared to non-variant carriers ⁽²⁸⁾. In young patients, the prevalence of germline variants in the BRCA2 gene is 2.9%, conferring a 23-fold greater risk of developing prostate cancer up to 56 years of age ⁽²⁸⁾.

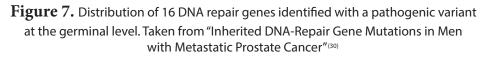


A la izquierda, se presenta la probabilidad de sobrevida en cáncer de próstata según si el paciente posee o no mutaciones germinales en el gen BRCA2 (p<0.0001). A la derecha, se presenta la probabilidad de sobrevida en cáncer de próstata en pacientes portadores de mutación en el gen BRCA2 según estadío de la enfermedad. Tomado de "The impact of a BRCA2 mutation on mortality from screendetected prostate cancer"⁽²⁹⁾.

In cases associated with Lynch syndrome, due to variants in the MMR genes involved in the repair of DNA replication errors, the risk of prostate cancer is three times higher compared to the general population⁽²⁷⁾.

In addition, germline variants have been reported in other DNA repair genes such as ATM, CHEK2, PALB2, RAD51D, and RAD51C, among others ⁽³⁰⁾.





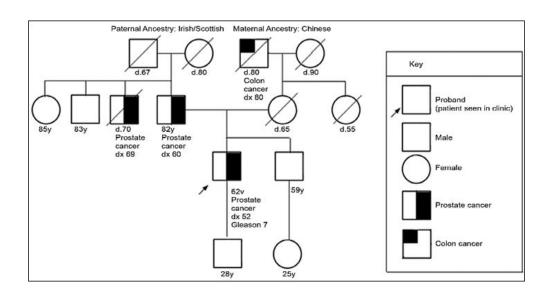


Figure 8. Family tree of a family with hereditary prostate cancer. The proband (arrow) and two relatives (first and second degree) have a diagnosis of prostate cancer. Taken from "How I Do It: Genetic counseling and genetic testing for inherited prostate cancer"⁽²⁷⁾

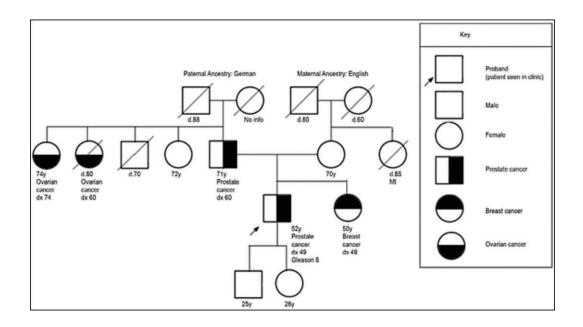


Figure 9. Family tree of a family with Hereditary Breast Ovarian Cancer Syndrome. The proband (arrow) with prostate cancer and two first-degree relatives (father and sister) were diagnosed with prostate cancer and breast cancer, respectively. He also has two second-degree relatives (paternal aunts) with ovarian cancer. Taken from "How I Do It: Genetic counseling and genetic testing for inherited prostate cancer" ⁽²⁷⁾

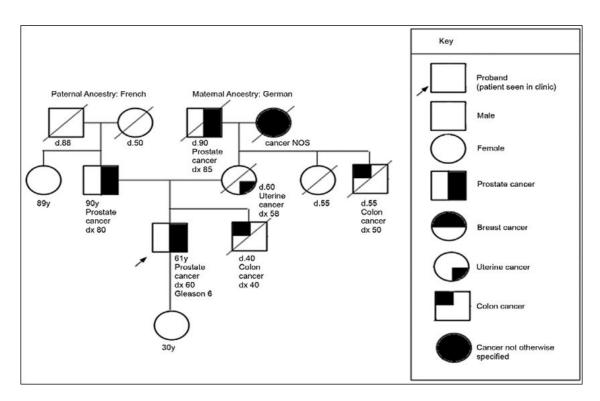


Figure 10. . Family tree of a family with Lynch syndrome. The proband (arrow) with prostate cancer and first and second relatives with neoplasms related to Lynch syndrome (colon, endometrial, and prostate cancer). Taken from "How I Do It: Genetic counseling and genetic testing for inherited prostate cancer"⁽²⁷⁾

CONCLUSION:

Knowing the molecular alterations of prostate cancer, as well as its molecular classification, genomic heterogeneity, and clonal evolution allows adequate management of patients. In addition, the collection of data from the patient's family history makes it possible

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to suspect a genetic predisposition to prostate cancer and thus refer him for timely genetic counseling and to be able to provide him with follow-up and therapeutic options.

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REFERENCES

- Khemlina G, Ikeda S, Kurzrock R. Molecular landscape of prostate cancer: implications for current clinical trials. Cancer Treat Rev. noviembre de 2015;41(9):761-6. <u>https://doi.org/10.1016/j.ctrv.2015.07.001</u>
- Demichelis F, Fall K, Perner S, Andrén O, Schmidt F, Setlur SR, et al. TMPRS52:ERG gene fusion associated with lethal prostate cancer in a watchful waiting cohort. Oncogene. 5 de julio de 2007;26(31):4596-9. https://doi.org/10.1038/sj.onc.1210237
- Sun X, Huang J, Homma T, Kita D, Klocker H, Schafer G, et al. Genetic alterations in the PI3K pathway in prostate cancer. Anticancer Res. mayo de 2009;29(5):1739-43. <u>https://ar.iiarjournals.org/content/29/5/1739.long</u>
- Taylor BS, Schultz N, Hieronymus H, Gopalan A, Xiao Y, Carver BS, et al. Integrative genomic profiling of human prostate cancer. Cancer Cell. 13 de julio de 2010;18(1):11-22. <u>https://doi.org/10.1016/j.ccr.2010.05.026</u>
- Robinson D, Van Allen EM, Wu Y-M, Schultz N, Lonigro RJ, Mosquera J-M, et al. Integrative clinical genomics of advanced prostate cancer. Cell. 21 de mayo de 2015;161(5):1215-28. <u>https://doi.org/10.1016/j.cell.2015.05.001</u>
- Netto GJ. Molecular Updates in Prostate Cancer. Surg Pathol Clin. diciembre de 2015;8(4):561-80. <u>https://doi.org/10.1016/j.path.2015.08.003</u>
- 7. Mitchell T, Neal DE. The genomic evolution of human prostate cancer. Br J Cancer. 14 de julio de 2015;113(2):193-8. https://doi.org/10.1038/bjc.2015.234
- Squire JA. TMPRSS2-ERG and PTEN loss in prostate cancer. Nat Genet. mayo de 2009;41(5):509-10. <u>https://doi.org/10.1038/ng0509-509</u>
- Bismar TA, Yoshimoto M, Vollmer RT, Duan Q, Firszt M, Corcos J, et al. PTEN genomic deletion is an early event associated with ERG gene rearrangements in prostate cancer: ERG REARRANGEMENTS AND PTEN DELETIONSIN PROSTATE CANCER. BUI Unt. febrero de 2011;107(3):477-85. https://doi.org/10.1111/j.1464-410X.2010.09470.x
- 10. Julia L Williams, Maisa Yoshimoto, Alexander H Boag, Jeremy A Squire, Paul C Park. TMPRSS2:ETS gene fusions in prostate cancer. http://atlasgeneticsoncology.org/Deep/TMPRSS2ERGinCancerID20091.ht ml.2010.
- 11. Cancer Genome Atlas Research Network. The Molecular Taxonomy of Primary Prostate Cancer. Cell. 5 de noviembre de 2015;163(4):1011-25. <u>https://doi.org/10.1016/j.cell.2015.10.025</u>
- Schoenborn JR, Nelson P, Fang M. Genomic profiling defines subtypes of prostate cancer with the potential for therapeutic stratification. Clin Cancer Res Off J Am Assoc Cancer Res. 1 de agosto de 2013;19(15):4058-66. <u>https://doi.org/10.1158/1078-0432.CCR-12-3606</u>
- Van Etten JL, Dehm SM. Clonal origin and spread of metastatic prostate cancer. Endocr Relat Cancer. abril de 2016;23(4):R207-217. <u>https://doi.org/10.1530/ERC-16-0049</u>
- Watson PA, Arora VK, Sawyers CL. Emerging mechanisms of resistance to androgen receptor inhibitors in prostate cancer. Nat Rev Cancer. diciembre de 2015;15(12):701-11. <u>https://doi.org/10.1038/nrc4016</u>
- 15. Wei L, Wang J, Lampert E, Schlanger S, DePriest AD, Hu Q, et al. Intratumoral and Intertumoral Genomic Heterogeneity of Multifocal Localized Prostate Cancer Impacts Molecular Classifications and Genomic Prognosticators. Eur Urol. 20 de julio de 2016; <u>https://doi.org/10.1016/j.eururo.2016.07.008</u>
- Boutros PC, Fraser M, van der Kwast T, Bristow RG. Clonality of localized and metastatic prostate cancer. Curr Opin Urol. mayo de 2016;26(3):219-24. <u>https://doi.org/10.1097/MOU.00000000000279</u>

- Beltran H, Yelensky R, Frampton GM, Park K, Downing SR, MacDonald TY, et al. Targeted next-generation sequencing of advanced prostate cancer identifies potential therapeutic targets and disease heterogeneity. Eur Urol. mayo de 2013;63(5):920-6. <u>https://doi.org/10.1016/j.eururo.2012.08.053</u>
- Beltran H, Demichelis F. Prostate cancer: Intrapatient heterogeneity in prostate cancer. Nat Rev Urol. agosto de 2015;12(8):430-1. <u>https://doi.org/10.1038/nrurol.2015.182</u>
- 19. Burrell RA, Swanton C. The evolution of the unstable cancer genome. Curr Opin Genet Dev. febrero de 2014;24:61-7. <u>https://doi.org/10.1016/j.gde.2013.11.011</u>
- Crea F, Nur Saidy NR, Collins CC, Wang Y. The epigenetic/noncoding origin of tumor dormancy. Trends Mol Med. abril de 2015;21(4):206-11. <u>https://doi.org/10.1016/j.molmed.2015.02.005</u>
- 21. Thalgott M, Rack B, Maurer T, Souvatzoglou M, Eiber M, Kreß V, et al. Detection of circulating tumor cells in different stages of prostate cancer. J C an cer R es C Clin O n col. m a yo de 2013;139(5):755-63. https://doi.org/10.1007/s00432-013-1377-5
- Merdan S, Tomlins SA, Barnett CL, Morgan TM, Montie JE, Wei JT, et al. Assessment of long-term outcomes associated with urinary prostate cancer antigen 3 and TMPRS52:ERG gene fusion at repeat biopsy. Cancer. 15 de noviembre de 2015;121(22):4071-9. <u>https://doi.org/10.1002/cncr.29611</u>
- Cary KC, Cooperberg MR. Biomarkers in prostate cancer surveillance and screening: past, present, and future. Ther Adv Urol. diciembre de 2013;5(6):318-29. <u>https://doi.org/10.1177/1756287213495915</u>
- 24. Biomarcadores en el cáncer de próstata. Implicación en la práctica clínica [Internet]. [citado 2 de agosto de 2016]. Disponible en: http://www.bvs.sld.cu/revistas/ibi/vol33_4_14/ibi11414.htm
- 25. Lin DW, Newcomb LF, Brown EC, Brooks JD, Carroll PR, Feng Z, et al. Urinary TMPRSS2:ERG and PCA3 in an active surveillance cohort: results from a baseline analysis in the Canary Prostate Active Surveillance Study. Clin Cancer Res Off J Am Assoc Cancer Res. 1 de mayo de 2013;19(9):2442-50.<u>https://doi.org/10.1158/1078-0432.CCR-12-3283</u>
- Brocks D, Assenov Y, Minner S, Bogatyrova O, Simon R, Koop C, et al. Intratumor DNA methylation heterogeneity reflects clonal evolution in aggressive prostate cancer. Cell Rep. 7 de agosto de 2014;8(3):798-806. https://doi.org/10.1016/j.celrep.2014.06.053
- Giri VN, Gross L, Gomella LG, Hyatt C. How I Do It: Genetic counseling and genetic testing for inherited prostate cancer. Can J Urol. abril de 2016;23(2):8247-53. <u>https://www.canjurol.com/abstract.php?ArticleID=&</u> <u>version=1.0&PMID=27085833</u>
- 28. Maia S, Cardoso M, Pinto P, Pinheiro M, Santos C, Peixoto A, et al. Identification of Two Novel HOXB13 Germline Mutations in Portuguese Prostate Cancer Patients. PloS One. 2015;10(7):e0132728. https://doi.org/10.1371/journal.pone.0132728
- 29. Akbari MR, Wallis CJD, Toi A, Trachtenberg J, Sun P, Narod SA, et al. The impact of a BRCA2 mutation on mortality from screen-detected prostate cancer. Br J Cancer. 9 de septiembre de 2014;111(6):1238-40. <u>https://doi.org/10.1038/bjc.2014.428</u>
- Pritchard CC, Mateo J, Walsh MF, De Sarkar N, Abida W, Beltran H, et al. Inherited DNA-Repair Gene Mutations in Men with Metastatic Prostate Cancer. N Engl J Med. 6 de julio de 2016. <u>https://doi.org/10.1056/NEJMoa1603144</u>

