



# CRITICAL VALUES FOR AUTOMATED BLOOD COUNTS AND PERIPHERAL BLOOD SLIDES

VALORES CRÍTICOS PARA HEMOGRAMAS AUTOMATIZADOS Y FROTIS DE SANGRE PERIFÉRICA

Huerto JL <sup>1a</sup>, Villaorduña AM <sup>1a</sup>

ORIGINAL PAPER

## ABSTRACT

**Objective:** The reporting of critical values is considered a necessary practice in clinical pathology laboratories since immediate communication determines therapeutic decisions that can save lives. The choice of critical values and the way they are reported must be a joint decision between laboratory specialists and clinical professionals from each health facility since they must be integrated into the dynamics of the services involved. Among the critical values that have been chosen for this review, we have included quantitative parameters, whose processing is carried out in automated hematology analyzers, and morphological findings in peripheral blood smears, which trained professionals evaluate.

**Keywords:** Pathology; Blood; Cytology; Laboratories; Microscopy. (Source: MESH-NLM)

## RESUMEN

**Objetivo:** El reporte de valores críticos se considera una práctica necesaria en los laboratorios de patología clínica, pues su comunicación inmediata determina decisiones terapéuticas que pueden salvar vidas. La elección de los valores críticos y el modo en que son reportados debe ser una decisión conjunta entre los especialistas del laboratorio y los profesionales clínicos de cada establecimiento de salud, ya que deben integrarse a la dinámica de los servicios involucrados. Entre los valores críticos que se han escogido para esta revisión hemos incluido parámetros cuantitativos, cuyo procesamiento se ejecuta en analizadores hematológicos automatizados, y hallazgos morfológicos en los frotis de sangre periférica, lo cuales son evaluados por profesionales capacitados.

**Palabras clave:** Patología; Sangre; Citología; Laboratorios; Microscopía. (Fuente: DeCS- BIREME)

<sup>1</sup> Servicio de Hematología, Hemoterapia y Banco de Sangre, Hospital Nacional Alberto, Sabogal Sologuren, Callao, Perú.

<sup>a</sup> Surgeon, specialty in clinical pathology.

Cite as: Huerto JL, Villaorduña AM. Critical values for automated blood counts and peripheral blood slides. Rev Fac Med Hum. 2022;22(4):697-706. doi 10.25176/RFMH.v22i4.4616

Journal home page: <http://revistas.urp.edu.pe/index.php/RFMH>

Article published by the Magazine of the Faculty of Human Medicine of the Ricardo Palma University. It is an open access article, distributed under the terms of the Creative Commons License: Creative Commons Attribution 4.0 International, CC BY 4.0 (<https://creativecommons.org/licenses/by/4.0/>), that allows non-commercial use, distribution and reproduction in any medium, provided that the original work is duly cited. For commercial use, please contact [revista.medicina@urp.edu.pe](mailto:revista.medicina@urp.edu.pe)



## INTRODUCTION

Critical values in clinical pathology are defined as results that reflect imminent threats to patients' lives unless therapeutic measures are administered promptly<sup>(1)</sup>. These results are often far outside the upper or lower limits of the "normal" ranges. Therefore, one should not fall into the error of considering "critical" any result that exceeds the reference values, that is, the expected values in healthy individuals<sup>(1)</sup>. The crucial difference between a critical and abnormal value is that, in the first case, the result indicates an urgent danger to the patient's life.

Likewise, a clear distinction must be made between critical and vital values. The latter are "values that represent a pathophysiological state so different from normal as to be life-threatening and for which corrective measures can be taken, but for which prompt action is not so crucial<sup>(2)</sup>." As we can see, a vital value is as important as a critical value, but the immediacy of its report is not so decisive.

Currently, clinical laboratories are responsible for determining and communicating their critical values, defined according to the complexity of each service, the type of users it serves, the resources available, and the local epidemiological profile. The effective practice of reporting critical values implies a decrease in mortality and morbidity in health systems and a reduction in costs derived from preventable complications and damages<sup>(3)</sup>.

In hematology laboratories, there is a lack of consensus about which results should be considered critical, as well as discrepancies in terminology and reporting mechanisms<sup>(4)</sup>. Although selecting critical values is the responsibility of each clinical pathology service, we consider it pertinent to propose a list of results (explaining, for each one, the reasons that justify their selection) that serves as a starting point for other laboratories to define or refine their results. own critical value reporting processes.

### CRITICAL VALUES FOR AUTOMATED BLOOD CELL COUNTS

#### Hemoglobin less than 7 g/dL

The mortality of postoperative patients who refused packed red cell transfusions for religious reasons has been investigated.

These studies have concluded that hemoglobin levels between 7 g/dL and 8 g/dL are associated with a low risk of mortality, while levels below 6 g/dL are associated with extremely high levels of mortality<sup>(5-7)</sup>. Bleeding (38.5%), respiratory failure (35.9%), kidney failure (28.2%), sepsis (20.5%), and cardiac infarction (12.8%) were the main causes of death for these patients<sup>(6)</sup>.

Likewise, the AABB's most recent clinical practice guideline (American Association of Blood Banks) recommends a hemoglobin threshold of 7 g/dL to indicate transfusion of packed red blood cells in hemodynamically stable patients<sup>(8)</sup>.

#### Hemoglobin greater than 22 g/dL and hematocrit greater than 65% in neonates

A hematocrit greater than 65% or a hemoglobin level greater than 22 g/dL define neonatal polycythemia: an abnormal elevation in the mass of circulating erythrocytes that generates hyperviscosity blood<sup>(9-11)</sup>. Hyperviscosity in neonates is associated with decreased blood supply to organs such as the brain, heart, lungs, and intestine. The resulting hypoperfusion can lead to life-threatening conditions such as systemic hypoxia, hypoglycemia, necrotizing enterocolitis, seizures, and other neurological symptoms<sup>(10)</sup>. Although the optimal management for neonates with polycythemia has not been standardized, exchange transfusion is usually indicated; however, this decision remains controversial<sup>(10-11)</sup>.

#### Hematocrit greater than 65%

In patients with cyanotic congenital heart disease, the main hematological complication consists of an increase in hematocrit and hemoglobin concentration. These changes are secondary to the production of erythropoietin in the kidneys, which, in turn, is induced by hypoxemia<sup>(12)</sup>. Erythropoietin stimulates the production of red blood cells in the bone marrow, causing erythrocytosis in peripheral blood.

This increase in the amount of circulating erythrocytes produces an increase in blood viscosity, which can condition stasis and blockage of the capillaries. Symptoms of hyperviscosity usually appear when the hematocrit exceeds a value of 65% and are especially related to a progressive increase in hematocrit<sup>(13)</sup>. Hyperviscosity syndrome, characterized by neurological and cardiopulmonary symptoms, can give rise to life-threatening complications, such as cardiac

infarction, thromboembolic events, and ischemia in multiple organs <sup>(14)</sup>.

### **Leukocyte count greater than 100,000/uL Hyperleukocytosis**

is defined as a leukocyte count greater than 100,000/uL <sup>(15-16)</sup>. This condition, in hematological pathologies such as acute myeloid leukemia and acute lymphoblastic leukemia, can produce a leukostasis syndrome. Leukostasis syndrome is characterized by neurological and cardiopulmonary symptoms and is considered a hemato-oncological emergency. In patients with acute myeloid leukemia, particularly, leukostasis is associated with a poor prognosis due to a high risk of early mortality and a high probability of long-term recurrence and death <sup>(16)</sup>. Rapid cytoreduction by leukapheresis has been proposed as a therapeutic action in patients with leukostasis due to acute leukemia; this measure has been shown to reduce early mortality in patients with leukostasis <sup>(17-18)</sup>.

### **Leukocyte count greater than 50,000/uL and a neutrophil count greater than 30,000/uL in neonates**

A persistent leukocytosis greater than 50,000/uL not associated with leukemia is called leukemoid reaction, its main causes being infections, intoxications, tumors solids, severe bleeding and hemolysis <sup>(19-20)</sup>. In neonates, the leukemoid reaction is defined by a white blood cell count greater than 50,000/uL or a neutrophil count greater than 30,000/uL and, in low birth weight patients, is associated with life-threatening conditions such as neonatal sepsis, intraventricular hemorrhage and bronchopulmonary dysplasia <sup>(21)</sup>. Mortality due to neonatal sepsis has been estimated between 11% and 19% <sup>(22)</sup>, and increases significantly in the case of multidrug-resistant microorganisms <sup>(23)</sup>. Likewise, in our environment, mortality due to intraventricular hemorrhage in low birth-weight neonates has been estimated at 47.1% <sup>(24)</sup>.

### **Neutrophil count less than 500/uL**

Febrile neutropenia, a condition defined by a neutrophil count less than 500/uL in the presence of fever (temperature greater than 38°C on two consecutive readings), is a significant cause of morbidity and mortality in cancer patients <sup>(25)</sup>. In adult cancer patients, in-hospital mortality associated with febrile

neutropenia has been estimated at 9.5%, although the percentages are significantly higher in patients with major comorbidities, fungal infections, and sepsis <sup>(26)</sup>. Likewise, in adult patients receiving myelosuppressive chemotherapy, febrile neutropenia is associated with a 15% higher mortality than in patients without febrile neutropenia <sup>(27)</sup>, since it predisposes a severe infection by fungi, gram-negative bacilli, and gram-positive cocci <sup>(28)</sup>.

In pediatric patients receiving myelosuppressive chemotherapy, febrile neutropenia is defined by a neutrophil count of less than 500 u/L or less than 1000/uL with a predicted decline over the next two days <sup>(29)</sup>. In this group of patients, in-hospital mortality associated with febrile neutropenia is estimated at 5% for gram-positive infections, 18% for gram-negative infections, and higher percentages for fungal infections <sup>(30)</sup>.

### **Platelet count less than 10,000/uL**

Thrombocytopenia is defined as a platelet count of less than 150,000/uL <sup>(31-32)</sup>. Patients with platelets above 50,000/uL often have no symptoms or clinical findings; bleeding from minimal trauma or prolonged bleeding from wounds usually appears with values below 30,000/uL and spontaneous bleeding events, considered a hematological emergency, occur with values below 10,000/uL <sup>(31-32)</sup>.

### **Platelet count greater than 1,000,000/uL**

A platelet value greater than 1,000,000/uL is associated with acquired von Willebrand syndrome: deficiency of von Willebrand factor multimers caused by increased proteolytic activity of ADAMTS13 <sup>(33)</sup>. The risk of bleeding is therefore increased in these extreme thrombocytoses. In patients with essential thrombocythemia, a platelet value greater than 1,000,000/uL, associated with a history of minor bleeding and a disease time greater than 15 years, is considered a high-risk factor for hemorrhage <sup>(34)</sup>. Likewise, it is recommended to moderate or restrict the use of aspirin in patients with essential thrombocythemia who reach platelet levels above 1,000,000/uL <sup>(33-34)</sup>.

**Table 1.** Critical values for automated blood counts.

CRITICAL VALUES FOR AUTOMATED BLOOD COUNTS		
PARAMETER	VALUES / FINDINGS	INTERPRETATION
Hemoglobin	< 7 g/dL	Risk of death in post-surgery. Recommended transfusion, with exceptions.
	> 22 g/dL	Neonates: Definition of polycythemia. Risk of hyperviscosity.
Hematocrit	> 65%	Hyperviscosity symptoms in heart patients. A possible indication of bleeding. Neonates: Definition of polycythemia. Exponential risk of hyperviscosity in neonatal polycythemia. Possible indication for exchange transfusion.
	> 100 000/uL > 50 000/uL (neonatos)	Risk of death due to leukostasis. Possible indication for leukapheresis. Risk of death in neonatal leukemoid reaction.
Neutrophils	< 500/uL	Risk of death associated with febrile neutropenia.
	> 30 000/uL (neonatos)	Risk of death in neonatal leukemoid reaction.
Platelets	< 10 000/uL	Risk of death from spontaneous bleeding.
	> 1 000 000/uL	Acquired von Willebrand factor deficiency, risk of bleeding, and aspirin contraindication.

## CRITICAL VALUES FOR PERIPHERAL BLOOD

### Smears

#### Schistocytes

The International Council for Standardization in Haematology (ICSH), which we can translate as the International Committee for Standardization in Hematology, indicates that a percentage of schistocytes greater than 1% of the total red blood cells, in the absence of other anomalies severe erythrocytes, has clinical importance for the diagnosis of microangiopathic hemolytic anemia<sup>(35)</sup>. Although a percentage of schistocytes greater than 1% can also be observed in renal failure, hemoglobinopathies, and neonates, it is usually accompanied by other erythrocyte morphological changes<sup>(36)</sup>.

One of the most frequent microangiopathic hemolytic anemias in adults, thrombotic thrombocytopenic purpura (TTP), has mortality close to 90% in the

absence of treatment<sup>(37)</sup>. With the introduction of therapeutic plasma exchange (TPR) in the 1980s, mortality was reduced to values close to 20%<sup>(38-39)</sup>. Due to the very high mortality of untreated TTP, TPR is considered to be indicated as an emergency procedure in these patients<sup>(39)</sup>.

#### Blasts

The finding of blasts in peripheral blood, by itself, does not imply an imminent risk to the life of patients; however, its identification and morphological description are crucial for making therapeutic decisions, and a delay in its reporting can predispose to potentially serious damage. For this reason, blasts fit the definition of "vital values": outcomes that are just as important as critical values, but for which immediate corrective action is not required<sup>(2)</sup>. Although there is no standardized recommendation for reporting vital

values, the need to include them as a logical extension of the critical values system has been recognized<sup>(40)</sup>.

The Classification of Hematopoietic and Lymphoid Tissue Tumors of the World Health Organization (WHO) establishes a value of 20% of blasts in peripheral blood or bone marrow to define acute leukemia<sup>(41)</sup>. Currently, however, we know that there is considerable intraindividual (the subject is compared against himself/herself at different times) and interindividual (the subject is compared against others) variability for microscopic examination and manual leukocyte count<sup>(42-43)</sup>. Because of this, a blast count of less than 20% may be "corrected" in a second reading that shows a percentage above this threshold. Therefore, even a percentage of blasts below 20% should be reported in patients without a history of leukemia, with the promptness corresponding to a vital value.

### Bacteria

Although the Wright stain, used in peripheral blood smears, is designed to identify leukocytes, red blood cells, and other cellular elements, it can also show microorganisms such as bacteria and parasites. The presence of bacteria in peripheral blood is a factor in poor prognosis and high mortality in patients with sepsis<sup>(44-45)</sup>. This is a very unusual finding, so possible contamination of the sample with bacteria from the central venous catheter must be ruled out<sup>(45)</sup>.

### Hemoparasites

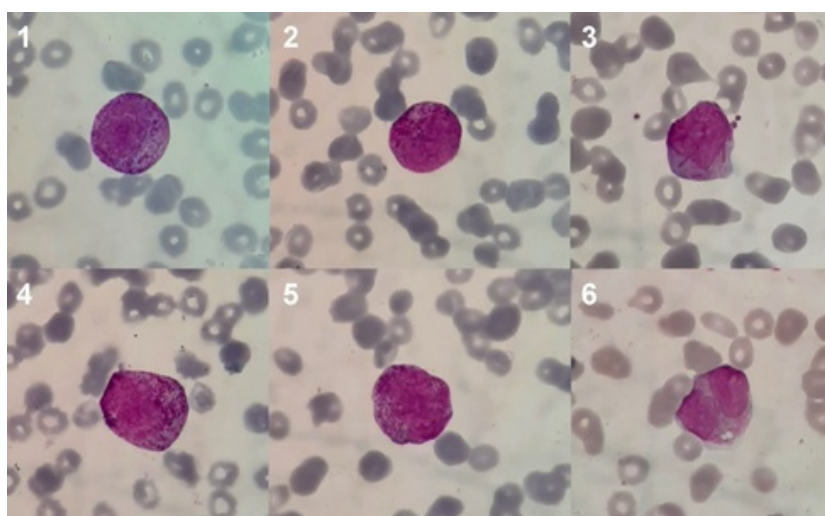
As in the case of blasts, a timely report of hemiparasites in peripheral blood is crucial for making therapeutic decisions; likewise, a delay in their identification could

lead to serious complications. For example, our country's timely malaria diagnosis is a public health priority. The WHO emphasizes the early diagnosis and treatment of malaria patients to reduce incidence and mortality rates<sup>(46)</sup>, a task in which clinical laboratories play a leading role. In this sense, a constant improvement has been verified, between 2012 and 2017, in the performance for the microscopic malaria diagnosis in the country's specialized laboratories<sup>(47)</sup>. In this context, Dr. Pedro Legua<sup>(48)</sup> highlights the importance of timely malaria diagnosis and stresses the need to identify *Plasmodium falciparum* correctly.

### Abnormal Promyelocytes

By "abnormal promyelocytes" (Figure 1) we mean promyelocytes with abnormal morphological characteristics that typically appear in the peripheral blood and bone marrow of patients with acute promyelocytic leukemia (APL). The WHO recognizes two morphological types of APL: the "classic" or hypergranular form and the microgranular or hypogranular variant<sup>(496)</sup>. Additionally, other morphological types with their characteristics have been described<sup>(50)</sup>.

Although the morphological presentation of APL is usually heterogeneous<sup>(50)</sup>, in all cases, it is associated with a high early mortality rate that approaches 17.5%<sup>(51)</sup>. This high mortality rate is related to the phenomenon of disseminated intravascular coagulation (DIC), which conditions events of hemorrhage and thrombosis<sup>(52)</sup>, with intracranial hemorrhage being the leading cause of death from ALI<sup>(53)</sup>. Therefore, starting treatment as soon as possible in patients with suspected ALI is recommended, even before the diagnosis is confirmed by molecular or genetic techniques<sup>(53)</sup>.



**Figure 1.** Anomalous promyelocytes in peripheral blood. <

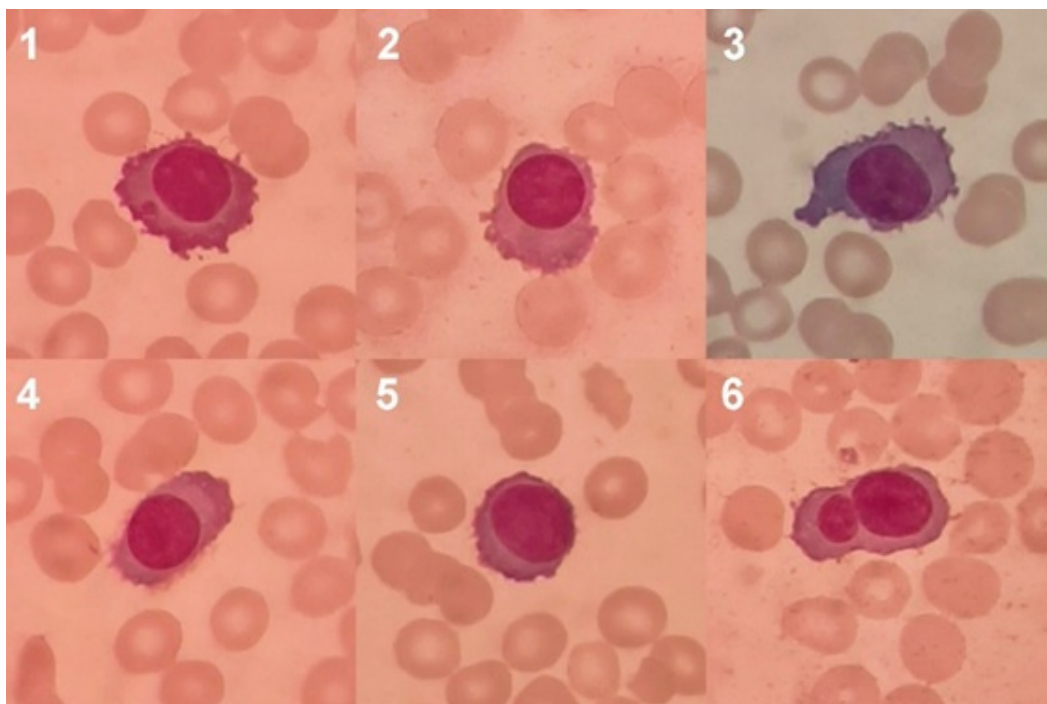
(Magnification: 100x. Coloration: Wright) Microphotographs 1-5: Anomalous promyelocytes of hypergranular morphological type. Photomicrograph 3: Hypergranular promyelocyte with multiple Auer bodies in the cytoplasm. Microphotograph 6: Anomalous promyelocyte of microgranular morphological type or "variant".

### Plasma cells

Plasma cells or plasmocytes (Figure 2) are lymphoid cells of the B lineage that originate in the bone marrow and whose function is to secrete antibodies. Although they constitute part of the normal cell population of the bone marrow, plasma cells are not usually found in the peripheral blood of healthy people<sup>(54)</sup>. As we will see, the finding of plasma cells in peripheral blood is related, in some patients, to a poor prognosis and high mortality.

In patients with multiple myeloma, the presence of circulating plasma cells in peripheral blood is associated with a poor clinical course, poor survival, and an increase in the International Staging System<sup>(55)</sup>.

Similarly, in patients with multiple myeloma, a percentage greater than 5% of plasma cells in peripheral blood is considered a marker of a highly proliferative disease, low life expectancy, and a prognosis similar to that of plasma cell leukemia<sup>(56)</sup>. Early mortality in plasma cell leukemia, although it has decreased in recent decades, is still significantly higher than in multiple myeloma<sup>(57)</sup>. This entity is defined by a plasma cell count greater than 20% of the total leukocytes in peripheral blood<sup>(58)</sup>, and it is considered the most aggressive plasma cell neoplasm with the worst prognosis<sup>(59)</sup>.



**Figure 2.** Plasma cells in peripheral blood.

(Magnification: 100x. Coloration: Wright) Photomicrographs 1-6: Plasma cells with intensely basophilic cytoplasm with an irregular border, fine cytoplasmic processes, clear perinuclear zone, round or oval nuclei with eccentric location and closed chromatin. Microphotograph 1: A cytoplasmic inclusion is observed. Microphotograph 5: Plasma cell with scant cytoplasm that could be mistakenly identified as an abnormal lymphocyte. Microphotograph 6: Binucleation is observed.

**Table 2.** Critical values for peripheral blood smears.

CRITICAL VALUES FOR PERIPHERAL BLOOD SMEARS		
PARAMETER	VALUES / FINDINGS	INTERPRETATION
Schistocytes	> 1% of total red blood cells	Risk of death from thrombotic thrombocytopenic purpura.
Blasts	20% of total leukocytes (consider reporting lower percentages, according to clinical-pathological correlation).	Immediate communication of the finding can prevent potentially serious damage.
Bacteria	Any amount.	Very poor prognosis in patients with sepsis.
Hemoparasites	Any number on a peripheral blood smear.	Immediate communication of the finding can prevent potentially serious damage.
Abnormal promyelocytes	Predominant cell type, lacking later stages of granulocytic maturation (myelocyte, metamyelocyte, neutrophil).	Risk of death from disseminated intravascular coagulation (DIC).
Correlation with other findings in the blood count and coagulation profile.		
Plasma cells	> 5% of total leukocytes.	Risk of death in multiple myeloma. A value above 20% is the diagnostic criteria for plasma cell leukemia, with an aggressive course and poor prognosis.

ORIGINAL PAPER

## DISCUSSION

One of the most important considerations for reporting critical values, in the particular case of the hematology laboratory, is that a correct and thorough morphological review be carried out reasonably. For proper identification of some cellular elements and abnormal shapes, an experienced observer must examine peripheral blood smears.

For example, regarding the presence of schistocytes, the ICSH establishes that the morphology is variable<sup>(35)</sup>: crescent-shaped schistocytes must be distinguished from sickle cells (sickle cells) based on their size, which in schistocytes is always smaller than a red blood cell; there are schistocytes with sharp angles or "spines" that look

like triangles; "helmet cells", damaged red blood cells with an "amputated" area recognizable by a rectilinear border, are considered to be equivalent to schistocytes; keratocytes, damaged red blood cells that appear to have a pair of "horns" separated by a concave segment, are also considered schistocyte equivalents; microspherocytes, likewise, should be included in the schistocyte count, but only in the presence of any of the previously mentioned forms.

The morphology of the abnormal promyelocytes of the LPA deserves a separate mention. Both morphological types mentioned by the WHO, the "typical" or hypergranular and the "variant" or microgranular, have



specific characteristics that differentiate the abnormal promyelocyte from its normal counterpart<sup>(49)</sup>.

Hypergranular promyelocytes have a cytoplasm filled with granules that range in color from pink to purple. These granules are compactly distributed in the cytoplasm and occasionally join or fuse (coalescence); they are usually comparatively larger than the granules of normal promyelocytes and may be so abundant they cover the nuclear rim, obscuring it. Other times, the granules are very fine and look like a "dust cloud" in the cytoplasm. In many cases, abnormal promyelocytes present multiple Auer bodies (rod-shaped, basophilic, crystalline cytoplasmic inclusions), which may adopt a bunch or bouquet distribution. The hypergranular promyelocyte nucleus is variable in size and shape, although it is usually bilobed or kidney-shaped<sup>(49-50)</sup>.

Microgranular promyelocytes, in contrast, are characterized by an apparent absence of cytoplasmic granules and a nucleus that is usually bilobed. Not infrequently, however, a small number of abnormal promyelocytes with distinguishable granules and clustered Auer bodies are also seen in microgranular APL. Currently, we know that the apparent lack of granules is due to the fact that they are submicroscopic in size and cannot be seen by light microscopy<sup>(49-50)</sup>.

Regarding plasma cells, since they are cells that are not normally observed in peripheral blood, special attention must be paid to correct morphological identification. The normal plasma cell is oval or circular, with abundant and intensely basophilic cytoplasm, a clear perinuclear zone corresponding to the Golgi apparatus, a low nucleus/cytoplasm ratio, and a small, eccentric nucleus with closed and lumpy chromatin. Additionally, in conditions such as multiple myeloma, plasma cells usually present atypical features in size, shape, and cytoplasmic coloration, different types of inclusions, abnormalities in the nuclear border, and multinucleation<sup>(60)</sup>.

On the other hand, more and more importance is

attached to the issuance of interpretive comments in laboratory tests; In our country, this responsibility corresponds to the medical specialist in clinical pathology. It is currently recommended that the laboratory results include a description or interpretation of the anomaly, information relevant to the diagnosis, suggestions for further examinations, and even an opinion related to the treatment<sup>(61)</sup>. Critical value reporting is considered an integral part of the interpretive reporting system, providing the clinician with information about life-threatening conditions and prompting prompt and decisive therapeutic action.

## CONCLUSIONS

The reporting of critical values is considered a necessary practice in laboratories and is beneficial in the clinical field. To have the desired impact, they need to be reported quickly, interpreted by a trained professional, and lead to prompt decision-making. Likewise, it is recommended that the critical values of each health establishment be chosen by consensus between laboratory professionals and those who work in clinical areas.

In the hematology laboratory, the critical values for complete blood counts comprise a series of quantitative parameters that, in most cases, are obtained directly from the automated hematology analyzer; in some cases, however, they can be corrected by the laboratory professional through a microscopic analysis (platelet count, leukocyte differential, etc.) or a manual procedure (microhematocrit by centrifugation, etc.). On peripheral blood smears, critical values include abnormal morphologic findings that often require evaluation by experienced professionals.

The recommendations proposed in this article can serve as a basis for other health establishments to prepare their own lists of critical values, considering that each laboratory service must do so in coordination with the interested clinical areas. Likewise, our recommendations can be used to expand the lists of critical values in those services that already have a notification system for these results.







**Authorship contributions:** the authors are the managers of the article in its entirety.

**Conflicts of interest:** There was no conflict of interest.

**Funding sources:** Self financed.

**Received:** may 22, 2022

**Approved:** august 28, 2022

**Correspondence:** José Luis Huerto.

**Address:** Jr. Colina 1081, Bellavista, Callao. Lima Perú.

**Telephone number:** 965266908

**E-mail:** joluahuag@gmail.com

## REFERENCES

- Campuzano Maya G. Valores críticos en el laboratorio clínico: de la teoría a la práctica. Med. Lab. [Internet]. 1 de julio de 2011 [citado 30 de julio de 2021];17(7-8):331-50. Disponible en: <https://medicinaylaboratorio.com/index.php/myl/article/view/362>
- Lundberg GD. Critical (Panic) Value Notification: An Established Laboratory Practice Policy (Parameter). JAMA [Internet]. 2 de febrero de 1990 [citado 30 de julio de 2021];263(5):709. Disponible en: <http://dx.doi.org/10.1001/jama.1990.03440050103044>
- Campuzano Maya G. La política de valores críticos es un derecho de los pacientes. Med. Lab. [Internet]. 1 de julio de 2011 [citado 30 de julio de 2021];17(7-8):309-10. Disponible en: <https://medicinaylaboratorio.com/index.php/myl/article/view/359>
- Keng TB, De La Salle B, Bourner G, Merino A, Han JY, Kawai Y et al. International Council for Standardization in Haematology (ICSH). Standardization of haematology critical results management in adults: an International Council for Standardization in Haematology, ICSH, survey and recommendations. Int J Lab Hematol. [Internet]. 18 de julio [citado 30 de julio de 2021]; 38(5):457-71. Disponible en: <https://onlinelibrary.wiley.com/doi/epdf/10.1111/ijlh.12526>
- Carson JL, Noveck H, Berlin JA, Gould SA. Mortality and morbidity in patients with very low postoperative Hb levels who decline blood transfusion. Transfusion. [Internet]. Julio de 2002 [citado 30 de julio de 2021];42(7):812-8. Disponible en: <https://onlinelibrary.wiley.com/doi/epdf/10.1046/j.1537-2995.2002.00123.x>
- Tobian AA, Ness PM, Noveck H, Carson JL. Time course and etiology of death in patients with severe anemia. Transfusion. [Internet]. Julio de 2009 [citado 30 de julio de 2021];49(7):1395-9. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/19389032/>
- Shander A, Javidrooz M, Naqvi S, Aregbeyen O, Caylan M, Demir S, Juhl A. An update on mortality and morbidity in patients with very low postoperative hemoglobin levels who decline blood transfusion (CME). Transfusion. [Internet]. Octubre de 2014 [citado 30 de julio de 2021];54:2688-95. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/27732721/>
- Carson JL, Guyatt G, Heddle NM, Grossman BJ, Cohn CS, Fung MK et al. Clinical Practice Guidelines From the AABB: Red Blood Cell Transfusion Thresholds and Storage. JAMA. [Internet]. 15 de noviembre de 2016 [citado 30 de julio de 2021];316(19):2025-2035. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/27732721/>
- Pantoja M. Policitemia neonatal e hiperviscosidad. Rev Soc Bol Ped. [Internet]. 2006 [citado 30 de julio de 2021]; 45 (1): 27 – 30. Disponible en: <http://www.scielo.org.bo/pdf/rbp/v45n1/v45n1a06.pdf>
- Remon J, Raghavan A, Maheshwari A. Polycythemia in the Newborn. NeoReviews. [Internet]. Enero de 2011 [citado 30 de julio de 2021]; 12(1): 20-28. Disponible en: <https://doi.org/10.1542/neo.12-1-e20>
- Alsafadi TR, Hashmi SM, Youssef HA, Suliman AK, Abbas HM, Albaloushi MH. Polycythemia in neonatal intensive care unit, risk factors, symptoms, pattern, and management controversy. J Clin Neonatol. [Internet]. Abril de 2014 [citado 30 de julio de 2021];3(2):93-8. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/25024975/>
- Peñuela O, Gómez L. Eritropoyetina: más allá de la proliferación y maduración eritroide. Rev. Fac. med. [Internet]. Junio de 2010 [citado 30 de julio de 2021];18(1): 67-76. Disponible en: <http://www.scielo.org.co/pdf/med/v18n1/v18n1a07.pdf>
- Ruiz J, García A. Síndrome hipoxémico crónico. Revista Española de Cardiología Suplementos. [Internet]. 2009 [citado 30 de julio de 2021]; 9(5): 13–22. Disponible en: <https://www.revespcardiolog.com/en-pdf-s1131358709732935>
- Perez Rogers A, Estes M. Hyperviscosity Syndrome. Treasure Island (FL): StatPearls Publishing. [Internet]. Enero de 2021 [citado 30 de julio de 2021]. Disponible en: <https://www.ncbi.nlm.nih.gov/books/NBK518963/?report=classic>
- Ali AM, Mirrakhimov AE, Abboud CN, Cashen AF. Leukostasis in adult acute hyperleukocytic leukemia: a clinician's digest. Hematol Oncol. [Internet]. Junio de 2016 [citado 30 de julio de 2021];34(2):69-78. Disponible en: <https://onlinelibrary.wiley.com/doi/epdf/10.1002/hon.2292>
- Röllig C, Ehninger G. How I treat hyperleukocytosis in acute myeloid leukemia. Blood. [Internet]. 21 de mayo de 2015 [citado 30 de julio de 2021];125(21):3246-52. Disponible en: <https://doi.org/10.1182/blood-2014-10-551507>
- Nan X, Qin Q, Gentile C, Ensor J, Leveque C, Pingali SR, Phan AT, Rice L, Iyer S. Leukapheresis reduces 4-week mortality in acute myeloid leukemia patients with hyperleukocytosis - a retrospective study from a tertiary center. Leuk Lymphoma. [Internet]. Setiembre de 2017 [citado 30 de julio de 2021];58(9):1-11. Disponible en: [https://www.uab.edu/medicine/pathology/images/On\\_Call\\_Schedules/Flyers\\_for\\_events/Qarmali\\_2.26.19\\_Leukapheresis\\_reduces\\_4\\_week\\_mortality\\_in\\_acute\\_myeloid\\_leukemia\\_patients\\_with\\_hyperleukocytosis\\_a\\_retrospective\\_study\\_from\\_a\\_tertiary\\_center.pdf](https://www.uab.edu/medicine/pathology/images/On_Call_Schedules/Flyers_for_events/Qarmali_2.26.19_Leukapheresis_reduces_4_week_mortality_in_acute_myeloid_leukemia_patients_with_hyperleukocytosis_a_retrospective_study_from_a_tertiary_center.pdf)
- Giammarco S, Chiusolo P, Piccirillo N, Di Giovanni A, Metafuni E, Laurenti L, Sica S, Pagano L. Hyperleukocytosis and leukostasis: management of a medical emergency. Expert Rev Hematol. [Internet]. Febrero de 2017 [citado 30 de julio de 2021];10(2):147-154. Disponible en: <https://doi.org/10.1080/17474086.2017.1270754>
- Sakka V, Tsioudras S, Giamarellos-Bourboulis EJ, Giamarellou H. An update on the etiology and diagnostic evaluation of a leukemoid reaction. Eur J Intern Med. [Internet]. Octubre de 2006 [citado 30 de julio de 2021];17(6):394-8. Disponible en: <https://www.ejinme.com/action/showPdf?pii=S0953-6205%2806%2900163-4>
- Granger JM, Kontoyiannis DP. Etiology and outcome of extreme leukocytosis in 758 nonhematologic cancer patients: a retrospective, single-institution study. Cancer. [Internet]. 1 de setiembre de 2009 [citado 30 de julio de 2021];115(17):3919-23. Disponible en: <https://acsjournals.onlinelibrary.wiley.com/doi/epdf/10.1002/cncr.24480>
- Duran R, Ozbek UV, Ciftdemir NA, Acunaş B, Süt N. The relationship between leukemoid reaction and perinatal morbidity, mortality, and chorioamnionitis in low birth weight infants. Int J Infect Dis. [Internet]. Noviembre de 2010 [citado 30 de julio de 2021]; 14(11): 998 - 1001. Disponible en: <https://www.ijidonline.com/action/showPdf?pii=S1201-9712%2810%2902455-0>
- Fleischmann-Struzek C, Goldfarb DM, Schlattmann P, Schlapbach LJ, Reinhart K, Kissoun N. The global burden of paediatric and neonatal sepsis: a systematic review. Lancet Respir Med. [Internet]. Marzo de 2018 [citado 30 de julio de 2021];6(3):223-230. Disponible en: [https://doi.org/10.1016/S2213-2600\(18\)30063-8](https://doi.org/10.1016/S2213-2600(18)30063-8)
- Wattal C, Kler N, Oberoi JK, Fursule A, Kumar A, Thakur A. Neonatal Sepsis: Mortality and Morbidity in Neonatal Sepsis due to Multidrug-Resistant (MDR) Organisms: Part 1. Indian J Pediatr. [Internet]. Febrero de 2020 [citado 30 de julio de 2021];87(2):117-121. Disponible en: <https://doi.org/10.1007/s12098-019-03106-z>
- Zea-Vera A, Turin CG, Rueda MS, Guillén-Pinto D, Medina-Alva P, Tori A et al. Intraventricular hemorrhage and periventricular leukomalacia in low birth-weight neonates in three hospitals in Lima, Peru. Rev Peru Med Exp Salud Publica. [Internet]. Julio de 2019 [citado 30 de julio de 2021];36(3):448-453. Disponible en: <http://www.scielo.org.pe/pdf/rins/v36n3/1726-4642-rins-36-03-448.pdf>
- Klastersky J, de Naurois J, Rolston K, Rapoport B, Maschmeyer G, Aapro M et al; ESMO Guidelines Committee. Management of febrile neutropenia: ESMO Clinical Practice Guidelines. Ann Oncol. [Internet]. Setiembre de 2016 [citado 30 de julio de 2021]; 27 (suppl 5): v111 - v118. Disponible en: <https://www.annalsofoncology.org/action/showPdf?pii=S0923-7534%2819%2931643-6>
- Kuderer NM, Dale DC, Crawford J, Cosler LE, Lyman GH. Mortality, morbidity, and cost associated with febrile neutropenia in adult cancer patients. Cancer. [Internet]. 15 de mayo de 2006 [citado 30 de julio de 2021];106(10):2258-66. Disponible en: <https://acsjournals.onlinelibrary.wiley.com/doi/epdf/10.1002/cncr.21847>
- Lyman GH, Michels SL, Reynolds MW, Barron R, Tomic KS, Yu J. Risk of mortality in patients with cancer who experience febrile neutropenia. Cancer. [Internet]. 1 de diciembre de 2010 [citado 30 de julio de 2021];116(23):5555-63. Disponible en: <https://acsjournals.onlinelibrary.wiley.com/doi/epdf/10.1002/cncr.25332>





28. Caggiano V, Weiss RV, Rickert TS, Linde-Zwirble WT. Incidence, cost, and mortality of neutropenia hospitalization associated with chemotherapy. *Cancer*. [Internet] 1 de mayo de 2005 [citado 30 de julio de 2021];103(9):1916-24. Disponible en: <https://acsjournals.onlinelibrary.wiley.com/doi/epdf/10.1002/cncr.20983>
29. Mendes AV, Sapolnik R, Mendonça N. New guidelines for the clinical management of febrile neutropenia and sepsis in pediatric oncology patients. *J Pediatr (Rio J)*. [Internet] Mayo de 2007 [citado 30 de julio de 2021];83(2 Suppl):S54-63. Disponible en: <https://www.scielo.br/j/jped/a/7F6Fs37t9VzdY4xbFbHt9w/?lang=en&format=pdf>
30. Davis K, Wilson S. Febrile neutropenia in paediatric oncology. *Paediatr Child Health (Oxford)*. [Internet] Marzo de 2020 [citado 30 de julio de 2021];30(3):93-97. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7172100/pdf/main.pdf>
31. Izak M, Bussel JB. Management of thrombocytopenia. *F1000Prime Rep*. [Internet] Junio de 2014 [citado 30 de julio de 2021];6:45. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4047949/pdf/medrep-06-45.pdf>
32. Gauer RL, Braun MM. Thrombocytopenia. *Am Fam Physician*. [Internet] 15 de marzo de 2012 [citado 30 de julio de 2021];85(6):612-22. Disponible en: <https://www.aafp.org/afp/2012/0315/afp20120315p612.pdf>
33. Falchi L, Bose P, Newberry KJ, Verstovsek S. Approach to patients with essential thrombocythaemia and very high platelet counts: what is the evidence for treatment? *Br J Haematol*. [Internet] Febrero de 2017 [citado 30 de julio de 2021];176(3):352-364. Disponible en: <https://onlinelibrary.wiley.com/doi/epdf/10.1111/bjh.14443>
34. Brière JB. Essential thrombocythemia. *Orphanet J Rare Dis*. [Internet] 8 de enero de 2007 [citado 30 de julio de 2021];2:3. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1781427/pdf/1750-1172-2-3.pdf>
35. Zini G, d'Onofrio G, Briggs C, Erber W, Jou JM, Lee SH, McFadden S, Vives-Corrons JL, Yutaka N, Lesesve JF; International Council for Standardization in Haematology (ICSH). ICSH recommendations for identification, diagnostic value, and quantitation of schistocytes. *Int J Lab Hematol*. [Internet] Abril de 2012 [citado 30 de julio de 2021];34(2):107-116. Disponible en: <https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1751-553X.2011.01380.x>
36. Schapkaiteiz E, Mezgebe MH. The Clinical Significance of Schistocytes: A Prospective Evaluation of the International Council for Standardization in Hematology Schistocyte Guidelines. *Turk J Haematol*. [Internet] 1 de marzo de 2017 [citado 30 de julio de 2021];34(1):59-63. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5451690/pdf/TJH-34-59.pdf>
37. Sánchez A. Actualización en Microangiopatías Trombóticas. *Hematología*. [Internet] Octubre de 2015 [citado 30 de julio de 2021];19:51-58. Disponible en: <http://www.sah.org.ar/revista/numeros/10-vol%2019-extraordinario.pdf>
38. Deford CC, Reese JA, Schwartz LH, Perdue JJ, Kremer Hovinga JA, Lämmle B, Terrell DR, Vesely SK, George JN. Multiple major morbidities and increased mortality during long-term follow-up after recovery from thrombotic thrombocytopenic purpura. *Blood*. [Internet] 19 de setiembre de 2013 [citado 30 de julio de 2021];122(12):2023-9. Disponible en: <http://www.jped.com.br/contendo/07-83-554/port.pdf>
39. George JN, Nester CM. Syndromes of thrombotic microangiopathy. *N Engl J Med*. [Internet] 14 de agosto de 2014 [citado 30 de julio de 2021];371(7):654-66. Disponible en: <https://www.nejm.org/doi/full/10.1056/nejma1312353>
40. Lundberg GD. It is time to extend the laboratory critical (panic) value system to include vital values. *MedGenMed*. [Internet] 29 de enero de 2007 [citado 30 de julio de 2021];9(1):20. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1925001/>
41. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, Bloomfield CD, Cazzola M, Vardiman JW. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood*. [Internet] 19 de mayo de 2016 [citado 30 de julio de 2021];127(20):2391-405. Disponible en: <https://reader.elsevier.com/reader/sd/pii/S0006497120301567?token=07A39C8684FFC8A3E97D8C22C780C94937C0CA814E0F4F18DE1EC4FAD7592101C77CCC88B3D31A311E0870058C3A8809&originRegion=us-east-1&originCreation=20210831201631>
42. Brambila E, Castillo-Guerra R y Lozano-Zarain P. Comparación entre tres métodos manuales empleados en la cuenta diferencial de leucocitos respecto a un equipo automatizado. *Bioquímica*. [Internet] Julio a Setiembre 2003 [citado 30 de julio de 2021];28(3):4-12. Disponible en: <https://www.redalyc.org/articulo.oa?id=57611572002>
43. Fuentes-Arderiu X, Dot-Bach D. Measurement uncertainty in manual differential leukocyte counting. *Clin Chem Lab Med*. [Internet] 2009 [citado 30 de julio de 2021];47(1):112-5. Disponible en: <https://doi.org/10.1515/CCLM.2009.014>
44. Gérard J, Lebas E, Godon A, Blanchet O, Geneviève F, Mercat A et al. Free and intracellular bacteria on peripheral blood smears: an uncommon situation related to an adverse prognosis. *Ann Biol Clin (Paris)*. [Internet] Enero y febrero de 2007 [citado 30 de julio de 2021];65(1):87-91. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/17264045/>
45. van der Meer W, Verwiel JM, Gidding CE, de Metz M, de Keijzer MH. Bacteria in blood smears: Overwhelming sepsis or trivial contamination. *Acta Haematol*. [Internet] 2002 [citado 30 de julio de 2021];107(4):220-3. Disponible en: <https://doi.org/10.1159/000058318>
46. Organización Mundial de la Salud. Paludismo. Puntos clave: Informe mundial sobre el paludismo 2017 [Internet]. Ginebra: Organización Mundial de la Salud; 2017 [citado el 27 de julio de 2021]. Disponible en: <http://www.who.int/malaria/media/worldmalaria-report-2017/es/>
47. Bartra-More C, Ramos-Castillo J, Mendoza-Bautista R, Galindo-Cabello N, Gebol-Cahuaza M, Chirinos-Palomino F. Evaluación del desempeño en el diagnóstico microscópico de malaria en la red de laboratorios del Instituto Nacional de Salud de Perú, 2012-2017. *Rev. Perú. med. exp. salud pública* [Internet]. 2019 [citado 30 de julio de 2021];36(3):469-474. Disponible en: <http://www.scielo.org.pe/pdf/rins/v36n3/1726-4642-rins-36-03-469.pdf>
48. Legua LP. Malaria en el Perú. *RMH* [Internet]. 28 de agosto de 2013 [citado 31 julio de 2021];5(3). Disponible en: <https://revistas.upch.edu.pe/index.php/RMH/article/view/436>
49. Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H et al. WHO classification of tumours of haematopoietic and lymphoid tissues. Revised 4th Edition. Lyon: International Agency for Research on Cancer; 2017. Disponible en: <https://ashpublications.org/blood/article/127/20/2375/35286/The-2016-revision-of-the-World-Health-Organization>
50. Neame PB, Soamboonsrup P, Leber B, Carter RF, Sunisloe L, Patterson W et al. Morphology of acute promyelocytic leukemia with cytogenetic or molecular evidence for the diagnosis: characterization of additional microgranular variants. *Am J Hematol*. [Internet] Noviembre de 1997 [citado 30 de julio de 2021];56(3):131-42. Disponible en: <https://onlinelibrary.wiley.com/doi/epdf/10.1002/%2851C1%291096-8652%28199711%2956%3A3%3C131%3A%3AAID-AJH1%3E3.0.CO%3B2-Z>
51. Park JH, Qiao B, Panageas KS, Schymura MJ, Jurcic JG, Rosenblatt TL, Altman JK, Douer D, Rowe JM, Tallman MS. Early death rate in acute promyelocytic leukemia remains high despite all-trans retinoic acid. *Blood*. [Internet] 4 de agosto de 2011 [citado 30 de julio de 2021];118(5):1248-54. Disponible en: <https://doi.org/10.1182/blood-2011-04-346437>
52. Fondevila CG. Coagulación intravascular diseminada. *Hematología*. [Internet] 2018 [citado el 30 de julio de 2021];22:37-43. Disponible en: [http://www.sah.org.ar/Revista/numeros/vol22/sup/09\\_Coagulacion\\_intravascular\\_diseminada.pdf](http://www.sah.org.ar/Revista/numeros/vol22/sup/09_Coagulacion_intravascular_diseminada.pdf)
53. Tallman MS, Lo-Coco F, Kwaan HC, Sanz MA, Gore SD. Early death in patients with acute promyelocytic leukemia. Proceedings from a live roundtable at the 2010 American Society of Hematology Annual Meeting, December 4-7, 2010, Orlando, Florida. *Clin Adv Hematol Oncol* [Internet]. Febrero de 2011 [citado el 30 de julio de 2021];9(2 Suppl 3):1-16. Disponible en: [https://www.hematologyandoncology.net/files/2013/04/ho0211\\_sup31.pdf](https://www.hematologyandoncology.net/files/2013/04/ho0211_sup31.pdf)
54. Wols HA. Plasma Cells. *Encyclopedia Life Science*. [Internet] junio de 2006 [citado 30 de julio de 2021]. Disponible en: <https://www.roitt.com/elspdf/Plasma.pdf>
55. Li J, Wang N, Tesfalual N, Gao X, Liu S, Yue B. Prognostic value of circulating plasma cells in patients with multiple myeloma: A meta-analysis. *PLoS One*. [Internet] 13 de julio de 2017 [citado 30 de julio de 2021];12(7):e0181447. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5509371/pdf/pone.0181447.pdf>
56. Granell M, Calvo X, García-Guiñón A, Escoda L, Abella E, Martínez CM et al. Prognostic impact of circulating plasma cells in patients with multiple myeloma: implications for plasma cell leukemia definition. *Haematologica*. [Internet] junio de 2017 [citado 30 de julio de 2021];102(6):1099-1104. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5451342/pdf/1021099.pdf>
57. Gonsalves WL, Rajkumar SV, Go RS, Dispenzieri A, Gupta V, Singh PP, Buadi FK et al. Trends in survival of patients with primary plasma cell leukemia: a population-based analysis. *Blood*. [Internet] 7 de agosto de 2014 [citado 30 de julio de 2021];124(6):907-12. Disponible en: <https://doi.org/10.1182/blood-2014-03-565051>
58. van de Donk NW, Lokhorst HM, Anderson KC, Richardson PG. How I treat plasma cell leukemia. *Blood*. [Internet] 20 de setiembre de 2012 [citado 30 de julio de 2021];120(12):2376-89. Disponible en: <https://ashpublications.org/blood/article/120/12/2376/30567/How-I-treat-plasma-cell-leukemia>
59. Wilson I, Gonsalves MD. Primary Plasma Cell Leukemia: A Practical Approach to Diagnosis and Clinical Management. *AJHO*. [Internet] 2017 [citado 30 de julio de 2021];13(3):21-25. Disponible en: [https://gotoper.com.s3.amazonaws.com/media/pdf/AJHO\\_Mar\\_Primary\\_cell\\_leukemia.pdf](https://gotoper.com.s3.amazonaws.com/media/pdf/AJHO_Mar_Primary_cell_leukemia.pdf)
60. Ribourtout B, Zandecki M. Plasma cell morphology in multiple myeloma and related disorders. *Morphologie*. [Internet] Junio de 2015 [citado 30 de julio de 2021];99(325):38-62. Disponible en: <https://doi.org/10.1016/j.morpho.2015.02.001>
61. Piva E, Plebani M. Interpretative reports and critical values. *Clin Chim Acta*. [Internet] Junio de 2009 [citado 30 de julio de 2021];404(1):52-8. Disponible en: <https://doi.org/10.1016/j.cca.2009.03.028>

